VOLUME 53, NUMBER 3, JULY, 1978

IDENTIFICATION OF SOME SYMPATHOMIMETIC AMINES BY THIN LAYER CHROMATOGRAPHY (TLC)

JITENDRA J. SHAH AND RAMILA J. SHAH

Woodson-Tenent Laboratories

Memphis, Tennessee 38101

ABSTRACT

The sympathomimetic amines were dissolved in water and ethanol, and treated with bromocresol green on a thin layer of alumina, silica gel and cellulose plates. Systems with two, three and four solvents were tested. The more polar spots were separated satisfactorily.

INTRODUCTION

Thin Layer Chromatography (TLC) is presently considered one of the most suitable techniques for drug analysis. It is fast, requires minimal equipment, and provides highly reliable results (Stahl, 1973). In addition, TLC is especially powerful because of its high sensitivity and resolution. Adequate separations of submicrogram quantities of material may be accomplished in under half an hour (Clarke, 1969).

In a previous report from this laboratory (Shah & Shah, 1976), sympathomimetic amines were separated satisfactorily using a two-solvent system. Rasin et al. (1969) showed that N-2,4 dinitro-phenylsulfenyl derivatives of some sympathomimetic amines are useful in the separation of the amines. The techniques used were ascending and multiple thin layer chromatography techniques on silica gel G or Al₂O₃ G. The best separa-

tion is obtained by multiple chromatography in 1:1 benzene-petroleum ether on silica gel G thin layers. The choices of solvent systems for the development of sympathomimetic amines are rather limited. Only systems containing either n-butanol or phenol are in practical use. These solvent systems are acidified either by acetic acid or by hydrochloric acid (Fabini & Konig, 1958a; Wachsmuth & Van Koeckhonen, 1962; Perry & Schroeder, 1963; Weidner & Weiss, 1958). Those most frequently used for paper chromatography (PC) are n-butanol, water, and acetic acid.

In TLC, the most commonly used sorbents are silica gel and cellulose; solvent systems are analogous to those for PC (Choulis, 1967a; DePotter et al., 1965). Sandri-Cavichi et al. (1966) used these systems for the separation of epinephrine and levarternol and for the separation of other adrenergic drugs as well. Halmekoski (1963) in some cases adds molybdate, tungstate or borate ions to the silica gel layer, an action which results in the marked decrease of mobility of compounds with two hydroxyl groups in the ortho position.

The purpose of this study was to develop systems and specific techniques suited for the identification of

some sympathomimetic amines. Chromatograms of five sympathomimetic amines have been developed with different organic solvents and water as the stationary phase. The amines were separated satisfactorily in the four solvent systems examined (Shah & Shah, 1976).

MATERIALS AND METHODS

Reagents

- (a) Solvents: Nanograde or spectrograde n-butanol, m-xylene, chloroform, ethanol and n-amyl alcohol (Fisher Scientific Co., 1241 Ambassador Blvd., St. Louis, MO 63132); ACS grade glacial acetic acid.
- (b) Bromocresol green: 0.01%. Dissolve 19 mg bromocresol green (Matheson, Colmen & Bell, Norwood, OH-45212, No. NB 179) in 100 ml ethanol.
- (c) Sympathomimetic amine standard solutions: Weigh 1 g of each amine (ephedrine hydrochloride, phenylpropanolamine hydrochloride, pyrilamine maleate, pheniramine maleate and chlorpheniramine maleate) into separate 100 ml volumetric flasks. Add 10 ml water and dilute to volume with ethanol (10 mg/ml).
- Apparatus (a) TLC plates: (1) Eastman Chromatogram Sheet, scribed silica gel without fluorescent indicator #13197 (scribed with 1 mm separations), 20 x 20 cm, 100 µm thick and cellulose without fluorescent indicator # 13255, 20 x 20 cm, 160 um thick (Eastman Kodak, Co., Rochester, NY 14650).
- (2) Precoated TLC plates, Uniplate alumina G, Avicel and silica gel G, 20 x 20 cm, 500 µm thick (Analab, Newark, DE 19711).
- (3) Precoated TLC plates, silica gel 60 and silica gel F-254 without fluorescent indicator, 20 x 20 cm, 0.25 mm layer thickness (EM Laboratories, Elmsford, NY 10523).
- (b) Developing tanks: Stainless steel or Glass, 9 x 9 x 4". (c) Ultraviolet (UV) light: Black-Ray UVL-22 (Ultraviolet Products, Inc., San Gabriel, CA 91778), 115 v, 60 cycles, equipped with short and long wavelengths.

Procedure

Apply 5 ul standard solutions side by side on alumina, cellulose or silica gel plates 25 cm from bottom and 40 mm apart (do not touch plate with pipet). Let spots dry. Apply 5 ul bromocresol green over each spot and let spots air dry. Amines are indicated by blue spots on greenish yellow background; spots will turn yellow in acetic acid vapors. Carry out chromatograms at room temperature until the solvent front has advanced 10 cm (30 to 45 min). Remove chromatograms from the solvent and dry in an oven at 100°C for ca 3 min. Examine derivatives spots (green-blue) under shortwave UV light. Determine R₁ value for each spot.

RESULTS AND DISCUSSION

The two main parameters involved in the identification of amines by TLC are related to detection and Rt values. Many reagents were tried on the amines bromocresol green, ninhydrine, eriochrom black-T, pnitrobenzoyl chloride and bromothymol blue. Only bromocresol green was found to react well with amines. The R_t values are only approximate values and are affected by many variables. Therefore, it is absolutely necessary to use several solvent systems and different modes of detection to identify amines.

A solution of these five sympathomimetic amines in ethanol showed one spot when treated with bromocresol green and chromatographed in the chloroform-ethanol solvent systems. The reagent itself gives one spot in this system. Table 1 indicates the Rr values of standards in the two-solvent systems. Kodak scribed silica gel was the best plate for the identification of amines in this system. Ephedrine hydrochloride, pyrilamine maleate and pheniramine maleate can be identified from the other amine standards.

A solution of these amines in ethanol showed one spot when treated with bromocresol green and chromatographed in the isopropanol-water-acetic acid and cyclohexane-benzene-diethylamine solvent systems. By using these systems, specific amines cannot be identified.

A solution of these amines in ethanol showed one and two spots when treated with bromocresol green and chromatographed in the n-butanol-m-xylene-wateracetic acid and n-amyl alcohol-m-xylene-water-acetic acid solvent systems, while the reagent gives only one spot. Table 2 indicates the Rr values of standards in the four-solvent systems. Uniplate avicel and Kodak cellulose were the best plates for the identification of amines in these system. Ephedrine hydrochloride, phenylpropanolamine hydrochloride, pyrilamine maleate and chlorpheniramine maleate can be identified in the latter system. Identification was made by comparison with amine standards.

TABLE 1: R₁ values of some sympathomimetic amines in two-solvent systems, after treatment with bromocresol green (45 minute developing time; all standards 10 mg/ml in ethanol).

Compound	So	olvent Sys	tem S ₁	S	olvent Syst	tem S2	Solvent System S ₃			
	Kodak		EM F-254	Kodak	EM 60	EM F-254	Kodak	EM 60	EM F-254	
Ephedrine hydrochloride	0.45	0.19	0.29	0.35	0.10	0.17	0.36	0.11	0.19	
Phenylpropanolamine hydrochloride	0.43	0.16	0.26	0.29	0.09	0.13	0.32	0.09	0.16	
Pyrilamine maleate	0.70	0.41	0.51	0.55	0.28	0.40	0.64	0.33	0.47	
Pheniramine maleate	0.60	0.22	0.31	0.53	0.17	0.24	0.55	0.19	0.29	
Chlorpheniramine maleate	0.62	0.23	0.32	0.50	0.20	0.26	0.56	0.21	0.31	
Reagent		0.07	0.08							

 $S_1 = \text{chloroform} + \text{ethanol} (80 + 20)$

 S_2 = chloroform + ethanol (83 + 17)

 S_3 = chloroform + ethanol (85 + 15)

Kodak = Kodak scribed silica gel

EM 60 = EM Laboratories silica gel 60

EM F-254 = EM Laboratories silica gel F-254

TABLE 2: Rt values of some sympathomimetic amines in four-solvent systems, after treatment with bromocresol green (30 minute developing time; all solutions mixed in 250 ml separatory funnel for ca. 5 minutes, aqueous layer discarded and organic layer used for TLC).

Solvent System S.			Solvent System S ₅			Solvent System S ₆				Solvent System S7					
Ks	U	Kc	EM	Ks	U	Kc	EM	Ks	U	Kc	EM	Ks	U	Kc	EM
0.23	0.42	0.42	0.14	0.31	0.67	0.53	0.18	0.18 0.27	0.43	0.48	0.14	0.30	0.63	0.56	0.18
0.18	0.30	0.32	0.16	0.35	0.45	0.41	0.20	0.07	0.32	0.36	0.15	0.35	0.43	0.43	0.21
0.06	0.82	0.70	0.09	0.19	0.91	0.77	0.13	0.07	0.80	0.78	0.09	0.18	0.88	0.85	0.13
0.06 0.14	0.79	0.68	0.07	0.19	0.89	0.75	0.13	0.07	0.78	0.77	0.06	0.17	0.86	0.82	0.11
0.11	0.85	0.79	0.10	0.22	0.94	0.84	0.13	0.17	0.86	0.88	0.09	0.22	0.92	0.92	0.13
0.05	0.16		0.07	0.12	0.29	0.20	0.13	0.07	0.17	0.18	0.08	0.11	0.26	0.27	0.13
	0.23 0.18 0.25 0.06 0.13 0.06 0.14 0.14	0.23 0.42 0.18 0.25 0.30 0.06 0.13 0.82 0.06 0.14 0.79 0.11 0.14 0.85 0.05	0.23 0.42 0.42 0.18 0.25 0.30 0.32 0.06 0.13 0.82 0.70 0.06 0.14 0.79 0.68 0.11 0.14 0.85 0.79 0.05	0.23 0.42 0.42 0.14 0.18 0.25 0.30 0.32 0.16 0.06 0.13 0.82 0.70 0.09 0.06 0.14 0.79 0.68 0.07 0.11 0.14 0.85 0.79 0.10 0.05	Ks U Kc EM Ks 0.23 0.42 0.42 0.14 0.31 0.18 0.25 0.30 0.32 0.16 0.35 0.06 0.13 0.82 0.70 0.09 0.19 0.06 0.14 0.79 0.68 0.07 0.19 0.11 0.14 0.85 0.79 0.10 0.22 0.05	Ks U Kc EM Ks U 0.23 0.42 0.42 0.14 0.31 0.67 0.18 0.25 0.30 0.32 0.16 0.35 0.45 0.06 0.13 0.82 0.70 0.09 0.19 0.91 0.06 0.14 0.79 0.68 0.07 0.19 0.89 0.11 0.14 0.85 0.79 0.10 0.22 0.94 0.05	Ks U Kc EM Ks U Kc 0.23 0.42 0.42 0.14 0.31 0.67 0.53 0.18 0.25 0.30 0.32 0.16 0.35 0.45 0.41 0.06 0.13 0.82 0.70 0.09 0.19 0.91 0.77 0.06 0.14 0.79 0.68 0.07 0.19 0.89 0.75 0.11 0.14 0.85 0.79 0.10 0.22 0.94 0.84 0.05	Ks U Kc EM Ks U Kc EM 0.23 0.42 0.42 0.14 0.31 0.67 0.53 0.18 0.18 0.25 0.30 0.32 0.16 0.35 0.45 0.41 0.20 0.06 0.13 0.82 0.70 0.09 0.19 0.91 0.77 0.13 0.06 0.14 0.79 0.68 0.07 0.19 0.89 0.75 0.13 0.11 0.14 0.85 0.79 0.10 0.22 0.94 0.84 0.13 0.05 0.05 0.12 0.20 0.20 0.13	Ks U Kc EM Ks U Kc EM Ks 0.23 0.42 0.42 0.14 0.31 0.67 0.53 0.18 0.27 0.18 0.25 0.30 0.32 0.16 0.35 0.45 0.41 0.20 0.11 0.06 0.13 0.82 0.70 0.09 0.19 0.91 0.77 0.13 0.11 0.06 0.14 0.79 0.68 0.07 0.19 0.89 0.75 0.13 0.15 0.11 0.14 0.85 0.79 0.10 0.22 0.94 0.84 0.13 0.26 0.05 0.05 0.07 0.12 0.20 0.20 0.12	Ks U Kc EM Ks U Kc EM Ks U 0.23 0.42 0.42 0.14 0.31 0.67 0.53 0.18 0.27 0.43 0.18 0.25 0.30 0.32 0.16 0.35 0.45 0.41 0.20 0.11 0.32 0.06 0.13 0.82 0.70 0.09 0.19 0.91 0.77 0.13 0.11 0.80 0.06 0.14 0.79 0.68 0.07 0.19 0.89 0.75 0.13 0.15 0.78 0.11 0.14 0.85 0.79 0.10 0.22 0.94 0.84 0.13 0.26 0.86 0.05 0.05 0.07 0.17 0.22 0.94 0.84 0.13 0.26 0.86	Ks U Kc EM Ks U Kc EM Ks U Kc 0.23 0.42 0.42 0.14 0.31 0.67 0.53 0.18 0.27 0.43 0.48 0.18 0.25 0.30 0.32 0.16 0.35 0.45 0.41 0.20 0.11 0.32 0.36 0.06 0.13 0.82 0.70 0.09 0.19 0.91 0.77 0.13 0.11 0.80 0.78 0.06 0.14 0.79 0.68 0.07 0.19 0.89 0.75 0.13 0.15 0.78 0.77 0.11 0.14 0.85 0.79 0.10 0.22 0.94 0.84 0.13 0.26 0.86 0.88 0.05	Ks U Kc EM Ks U Kc EM Ks U Kc EM 0.23 0.42 0.42 0.14 0.31 0.67 0.53 0.18 0.27 0.43 0.48 0.14 0.18 0.27 0.30 0.32 0.16 0.35 0.45 0.41 0.20 0.11 0.32 0.36 0.15 0.06 0.13 0.82 0.70 0.09 0.19 0.91 0.77 0.13 0.11 0.80 0.78 0.09 0.06 0.14 0.79 0.68 0.07 0.19 0.89 0.75 0.13 0.15 0.78 0.77 0.06 0.11 0.85 0.79 0.10 0.22 0.94 0.84 0.13 0.26 0.86 0.88 0.09 0.05 0.05 0.07 0.13 0.26 0.86 0.88 0.09	Ks U Kc EM Ks 0.23 0.42 0.42 0.14 0.31 0.67 0.53 0.18 0.27 0.43 0.48 0.14 0.30 0.18 0.25 0.30 0.32 0.16 0.35 0.45 0.41 0.20 0.11 0.32 0.36 0.15 0.35 0.06 0.13 0.82 0.70 0.09 0.19 0.91 0.77 0.13 0.11 0.80 0.78 0.09 0.18 0.06 0.14 0.79 0.68 0.07 0.19 0.89 0.75 0.13 0.15 0.78	Ks U Kc EM Ks U Xs U Xs U Xs U Xs U Xs U Xs	Ks U Kc EM Ks U Ks U Kc EM Ks U Ks

 $S_0 = \text{n-amyl alcohol} + \text{m-xylene} + \text{water} + \text{acetic acid} (15 + 85 + 10 + 40)$ Kc = Kodak cellulose EM = EM Laboratories silica gel F-254 $S_1 = n$ -amyl alchohol + m-xylene + water + acetic acid (20 + 80 + 10 + 40)

CONCLUSION

The more polar spots have Rr values which are approximately the same, and the less polar spots have different Rr values, thus producing separation. By using two- and four-solvent systems, we can identify some amines in the presence of others.

ACKNOWLEDGMENT

The authors acknowledge with thanks the use of research facilities at the Woodson-Tenent Laboratories, Inc., to undertake these investigations.

LITERATURE CITED

Choulis, N. H. 1967a. Separation and Quantitative Determination of Ardenaline using TLC. J. Pharm. Sci. 56:196.

Clarke, E. G. C. 1969. Isolation and Identification of Drugs, The Pharmaceutical Press, 17, Bloomsbury Square, WCI, London, 43 p.

DePotter, W. P.; Vochten, R. F. et al. 1965. TLC of catecholamines and their metabolites.

Halmekoski, J. 1963. TLC of adrenaline derivatives on layers containing chelate forming anions. Sumeon Kemistilehti, B, Perry, T. L. and Schroeder, W. A. 1963. The occurrence of amines in human urine, Determination by combined ion exchange and PC. J. Chromatography. 12:358.

Pohloudek-Fabini, R. and Konig, K. 1958a. PC behavior of the sympathomimetic substances. I. contribution to the analysis of several sympathomimetic substances. Pharmazie.

Rasim, T. and Zeytinoglu, L. 1969. 2,4 dinitrophenyl sulfenyl derivatives of sympathomimetic amines II TLC. Istanbul Uni Eszacilik Fak Mecm. 5(3):159.

Sandri-Cavicchi, G.; Quaglio, M. P. and Braghirolli, A. 1966. Separation of sympathomimetic agents by TLC. Boll. Chim. Farm. 105:670.

Shah, J. J. and Shah, R. J. 1976. TLC identification of some sympathomimetic amines. J. AOAC. 59:1416.

Shah, J. J. and Shah, R. J. 1976. TLC identification of some sympathomimetic amines. 90th Annual Meeting of the AOAC, Octo 1976, Washington, D.C. #48.

Stahl, E. 1973. "Drug analysis by chromtography and Microscopy". 1st edition, Ann Arbor Science Publishers, Ann Arbor, Mich. 3 p.

Wachsmuth, H. and Van Koeckhonen, L. 1962. Determination of sympathol and phenylephrine. J. Pharm. Belg. 17:227.

Weidner, G. and Weiss, F. 1958. Detection of adrenaline by PC and in the presence of other phenylethyl derivatives. Pharmazie, 13: 395.