# VARIATION IN LINEAR GROWTH AND SPORULATION OF SINGLE SPORE ISOLATES OF HELMINTHOSPORIUM MAYDIS

P. P. Hunter<sup>1</sup> and J. W. HILTY<sup>2</sup>
University of Tennessee, Knoxville, Tennessee 37901

#### ABSTRACT

Single spore isolates of *Helminthosporium maydis* exhibited variation in linear growth and sporulation when cultured on corn meal agar, lactose casein hydrolysate medium, potato-dextrose agar and V-8 juice. Also, the media differed in supporting growth and inducing sporulation.

### Introduction

Following the 1970 epiphytotic of southern corn leaf blight, race T of the pathogen, *Helminthosporium maydis* Nisikado and Miyake, was designated. This new race is host specific for corn lines produced with Texas (T) male-sterile cytoplasm. The studies presented in this paper were undertaken to assess the variation in sporulation and linear growth that exists in the *H. maydis* population.

Several studies have dealt with the effect of nutrition on growth and sporulation of Helminthosporium spp. Malca and Ullstrup (1972), working with H. carbonum Ullstrup and H. turcicum Pass, found variation in sporulation greater than variation in mycelial growth in response to several carbon (C) and nitrogen (N) sources. Maloul (1960) determined that for H. carbonum and H. turcicum the C source in the medium affected which N source was used most effectively in spore production, and the amount and rate of growth determined to a large extent the factors which favored sporulation. The work of Kafi and Tarr (1966), using five species of Helminthosporium, indicated that sporulation occurred over a lower range of C/N ratios than did vegetative growth. Reddy (1970) showed that lactose was poorly utilized by five species of Helminthosporlum, but utilization was improved when half of the lactose was replaced by its hydrolytic components. Zaman and Ahmed (1971i varied the sugar content of a basic medium and found that mycelial growth of H. oryzae Breda de Haan increased with an increase in sugar content up to 3%. Studying H. cynodontis Margi., White and Johnson (1971) found that linear growth was increased by trace amounts of zinc.

## MATERIALS AND METHODS

Cultural variability in linear growth and sporulation among forty-two single spore isolates of Helminhosporium maydis was studied using four cultural media. The six cultures from each of which seven single spore isolates were obtained were originally isolated from lesions on corn. The six original cultures are referred to as sources in this paper. The four media used were: corn meal agar (CMA) (Johnson, 1972), lactose casein

hydrolysate medium (LCHA) (Tuite, 1969), potato-dextrose agar (PDA) (Tuite, 1969), and V-8 juice agar (VJA) (Tuite, 1969). Isolates were maintained on PDA previous to the test, A 7-mm cork borer was used to cut uniform discs from the margins of cultures. These discs were transferred to the centers of five 90-mm test plates. Plates were numbered and randomized within a stack and the stacks were randomized in an incubator. Percival E-54U environmental chambers served as incubators and were maintained in complete darkness at 26 ± 2°C throughout the test. Each plate served as one replication. After four days incubation, the largest colony diameter on each of three plates was measured and the plates and stacks were rerandomized. After 48 hours, two plates, each serving as one replication, were chosen at random from each cultural source for spore counts. Spore suspensions were obtained by flooding the plates with a 5% sucrose - 0.2% "Tween 20" solution and scraping the surface of the colony. The resulting spore suspension was poured through cheesecloth into a beaker and the filtrate volume adjusted to 50 ml with the suspending solution. Spore suspensions were agitated, and a sample transferred to a hymacytometer slide. Four 1mm8 grids were counted and averaged to give a spore concentration for each plate. These values were expressed as spores per milliliter and were adjusted for colony growth by dividing by the colony area to obtain the number of spores/mm<sup>2</sup>.

## RESULTS AND DISCUSSIONS

The range and mean values for linear growth and sporulation are presented in Table 1. The means in Table 1 are overall means for all isolates. Linear growth on LCHA, CMA, and VJA was not significantly less than on the other three media.

The degree of variability on a given medium can be used as a measure of the extent to which that medium distinguishes differences among a group of isolates. The level of probability values for the analysis of variance tests among the 42 single spore isolates for each medium are a measure of the degree of variability on each medium. These values for both linear growth and sporulation are presented in Table 2. There were no differences among the four media in distinguishing variation in linear growth. Greatest differences in sporulation among the single spore isolates were obtained on PDA and LCHA. There were no significant differences in sporulation on CMA or VJA. The linear growth data for the 42 single spore isolates by sources on each media is presented in Table 3. Ten single spore isolates from five of the sources had significantly different linear growth among the four media. The sporulation data for the 42 single spore isolates by sources on each media is presented in Table 3. Ten single spore isolates from five of the sources had significantly different linear growth among the four media. The sporulation data for the 42 single spore isolates by sources on each media is presented in Table 4. Fifteen single spore isolates from the six sources had significant differences in sporulation among the four media. Comparison of Tables 3 and 4 will show that

three single spore isolates (G7, X4 and X6) had significant differences in linear growth and sporulation on the four media.

Single spore isolates from two of the sources had extremely low spore production while maintained on PDA. Tuite (1969) lists lactose casein hydrolysate medium as a good medium for inducing sporulation in Helminthosporium spp. The isolates from these sources

#### TABLE 1:

Range and mean values for linear growth and sporulation of 42 single spore isolates of Helminthosporium maydis on corn mean agar (CMA), lactose casein hydrolysate medium (LCHA), potato-dextrose agar (PDA), and V-8 juice agar (VJA)

	Linear Growth (mm) 1			Sporulation (Spores/mm <sup>2</sup> x 1000) <sup>2</sup>	
Media	Range	Mean	Media	Range	Hean
LCHA	78.00-31.00	59.61a <sup>3</sup>	PDA	9.180-0.000	2.893a
CHA	74.33-40.67	58.26a	LCHA	3.965-0.000	1.5856
VJA	80.67-36.00	58.20a	VJA	3.910-0.000	1.376b
PDA	71.67-20.00	52.96Ъ	CHA	2.565-0.000	0.271c

 $<sup>^{\</sup>mathrm{I}}\mathrm{Values}$  are means for three replications of all 42 isolates.

TABLE 2: Level of probability values for corn meal agar (CMA), lactose casein hydrolysate medium (LCHA), potato-dextrose agar (PDA), and V-8 juice agar (VJA) for linear growth and sporulation

Media	Sporulation	Linear Growth
CMA	5.29	0.01**
LCHA	0.15 **	0.01**
PDA	0.01 **	0.01**
VJA	4.94	0.01**

had sporulation values on LCHA significantly greater than on the other three media, These results substantiate that lactose casein hydrolysate medium is a good medium for inducing sporulation in *Helminthosporium maydis*.

All the isolates which sporulated sufficiently in culture were determined to be *H. maydis* race T in greenhouse inoculation tests of corn inbreds with the Texas (T) male-sterile and normal (N) cytoplasm.

TABLE 3: Linear growth of 42 single spore isolates of Helminthosporium maydis on four media—corn meal agar (CMA), lactose casein hydrolysate medium (LCHA), potato-dextrose agar (PDA), and V-8 juice agar (VJA)

11 56.00a <sup>2</sup> 62.67a 61.33a 62. 2 62.33a 65.67a 66.67a 65. 3 76.00a 78.00a 69.33a 73. 4 71.67a 72.33a 71.67a 74. 5 66.67a 63.00a 38.67b 68. 6 50.67a 63.00a 38.67b 68. 6 50.67a 65.50a 37.67a 49. 7 66.67a 61.00a 64.33a 48. 21 52.67a 31.00a 38.33a 36. 2 45.00a 52.33a 52.33a 49. 3 47.67a 52.33a 52.33a 49. 3 47.67a 52.33a 52.33a 52.33a 49. 4 46.67c 46.67c 52.33b 58. 6 56.67a 67.00a 66.00a 62. 6 56.67a 67.00a 66.00a 62. 6 56.67a 31.00a 38.33a 36. 7 47.33a 59.67a 44.33a 34. 81 50.67a 31.00a 38.33a 36. 81 50.67a 31.00a 52.33a 52.33a 52.33a 52.33a 52.33a 52.33a 52.33a 52.33a 53. 81 50.67a 67.00a 66.00a 62. 81 50.67a 59.67c 48.67b 46. 81 50.67a 31.00a 38.33a 36.00a 55.1 81 50.67a 31.00a 38.33a 36.00a 55.1 81 50.67a 31.00a 38.33a 36.00a 55.1 81 50.67a 59.67c 48.67b 46. 81 50.67a 59.67c 48.67b 46. 81 50.67a 59.67c 48.67b 46. 81 50.67a 59.03a 57.00a 41. 81 50.67a 59.33a 56.00a 53.47 59.1 81 50.67a 59.33a 67.33a 6					
11 56.00a <sup>2</sup> 62.67a 61.33a 62. 2 62,33a 65.67a 66.67a 65. 3 76.00a 78.00a 69.33a 73. 4 71.67a 72.33a 71.67a 74. 5 66.67a 63.00a 38.67b 68. 6 50.67a 55.00a 57.67a 49. 7 65.67a 61.00a 64.33a 48. 21 52.67a 31.00a 38.33a 36. 2 45.00a 52.33a 52.33a 49. 3 47.67a 52.33a 57.67a 54. 4 46.67c 46.67c 52.33b 58. 5 58.67a 67.00a 66.00a 62. 6 56.67a 65.67a 59.00a 58. 81 50.67a 31.00a 38.33a 36. 81 50.67a 31.00a 58.33a 57.67a 54. 6 56.67a 67.00a 66.00a 62. 6 56.67a 67.00a 66.00a 62. 6 56.67a 59.67c 44.33a 54. 81 50.67a 31.00a 38.33a 36.00a 55. 7 47.33a 59.67a 44.33a 54. 81 50.67a 31.00a 38.33a 36.00a 55. 6 56.33a 49.33a 36.00a 55. 7 55.63a 59.67a 59.00a 67.33a 56.00a 55. 7 55.67a 55.00a 67.33a 57.05a 59.06 66.05a 62.33a 58.33a 36.7b 64. 8 59.00b 62.33ab 79.33a 73.67b 64. 8 59.00b 62.33ab 79.33a 73.67b 64. 8 59.00b 62.33ab 79.33a 73.67b 64. 8 59.00b 62.33ab 79.33a 67.33a 67.35a 67. 7 55.67a 55.00a 67.33a 73.36 68.00a 69.00a 60.00a 65.00a 60.00a 66.00a 62.33a 73.36 64.67a 59.33a 67.33a 67.33			Growth	(mm) <sup>1</sup>	
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3 66.67a 69.67a 64.67a 75. 4 76.33a 73.67a 65.67b 64. 5 62.33a 62.00a 47.33a 48. 6 70.00a 57.33b 41.33c 71. 7 61.33a 69.33a 66.00a 68	2 3 4 5 6 7	75.00a 66.67a 76.33a 62.33a 70.00a 61.33a	71.67a 69.67a 73.67a 62.00a 57.33b 69.33a	41.33b 64.67a 65.67b 47.33a 41.33c 66.00a	69.00a 72.67a 75.00a 64.00b 48.00a 71.00a 68.67a 80.67a

lyalues are means of three replications.

<sup>1</sup> Present address: Department of Plant Pathology and Physiology, Virginia Polytechnic Institute and State University, Blacksburg, VA.

<sup>&</sup>lt;sup>2</sup> Associate Professor of Plant Pathology, Department of Agricultural Biology, University of Tennessee, Knoxville, TN.

<sup>&</sup>lt;sup>2</sup>Values are means for two replications of all 42 isolates.

<sup>.3</sup> Heans followed by the same letter are not significantly different at the 5% level of probability as determined by Duncan's multiple range test.

<sup>&</sup>lt;sup>2</sup>Means followed by the same letter are not significantly different at the 5% level of probability as determined by Duncan's multiple range test.

TABLE 4: Sporulation of 42 single spore isolates of Helminthosporium maydis on four media—corn meal agar (CMA), lactose casein hydrolysate medium (LCHA), potato-dextrose agar (PDA), and V-8 juice agar (VJA)

	Sporulation (Spores/mm <sup>2</sup> X 1000) <sup>1</sup>				
Isolate	CMA	LCHA	PDA	VJA	
11	0.105b <sup>2</sup>	3.865a	0.000ь	0.000ь	
2	0.000a	0.000a	0.000a	0.000a	
3	0,000a	0.000a	0.000a	0.000a	
4	1,620a	0.000a	0.000a	0.000a	
5	0.000a	0.000a	0.000a	0.000a	
6	0.000b	2.320a	0.770ь	0.450b	
7	0.000a	0.000a	0.000a	0.000a	
Z1	0.000a	3.360a	0.355a	1.000a	
2	0.070b	3.905a	0.895b	0.645b	
3	0.065a	3.680a	0.055a	1.055a	
5	0.000a	3.680a	0.000a	0.000a	
5	0.060b	1.715a	0.495b	1.515a	
6	0.000a	1.215a	0.065a	0.100a	
7	0.000a	0.000a	0.000a	0.000a	
R1	0.185b	3.245b	8.370a	2.255b	
2	0.595a	0.315a	4.340a	2.065a	
3	0.285a	0.920a	0.000a	0.645a	
4 5 6	0.120c	1.055bc	3.145a	1.605b	
5	0.865a	2.850a	6.890a	2.860a	
	0.235a	1.155a	3.680a	1.075a	
7	2.565a	3.965a	9.180a	1.185a	
Cl	0.715a	1.540a	2.580a	2.435a	
2	0.500b	1.190ab	1.950a	1.765a	
4	0.125a	0.960a	3.100a	1.955a	
5 6 7	0.210c	2.195ab	2.905a	1.375bc	
6	0.730a	1.305a	6.205a	2.395a	
7	0.000ь	1.135ab	3.145a	2.360ab	
61	0.315a	0.820a	4.160a	2.605a	
2	0.225a	1.520a	3.445a	1.505a	
3	0.200a	0.690a	3.405a	1.365a	
4	0.340c	0.685c	3.165a	2.190ь	
5	0.535a	1.200a	5.345a	3.910a	
6	0.260a	0.930a	2.530a	2.215a	
7	0.150b	0.460b	2.075a	0.060b	

TABLE 4: (continued)

Isolate	Sporulation (Spores/mm <sup>2</sup> X 1000)1				
	CMA	LCHA	PDA	VJA	
X1	0.335c <sup>2</sup>	0.665bc	3.545a		
2	0.060a	1.925a	7.665a	1.7156	
3	0.340c	1.845b	3.945a	2.145a	
4	0.280c	1.525bc	5.010a	1.4756	
5	0.100c	2.900b	7.085a	2.465b 1.835b	
6	0.195b	1.2806	6.795a	1.8606	
7	0.390a	7.840a	2.745a	1.620a	
8	2.485a	2.230a	1.790a	0.245a	

lValues are means of two replications.

<sup>2</sup>Means followed by the same letter are not significantly different at the 5% level of probability as determined by Duncan's multiple range test.

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